

# MSc Research Opportunities

in the Microbial Ecology & Technology-Lab  
(**METlab**) **Nov 2015**

**Barth F. Smets**

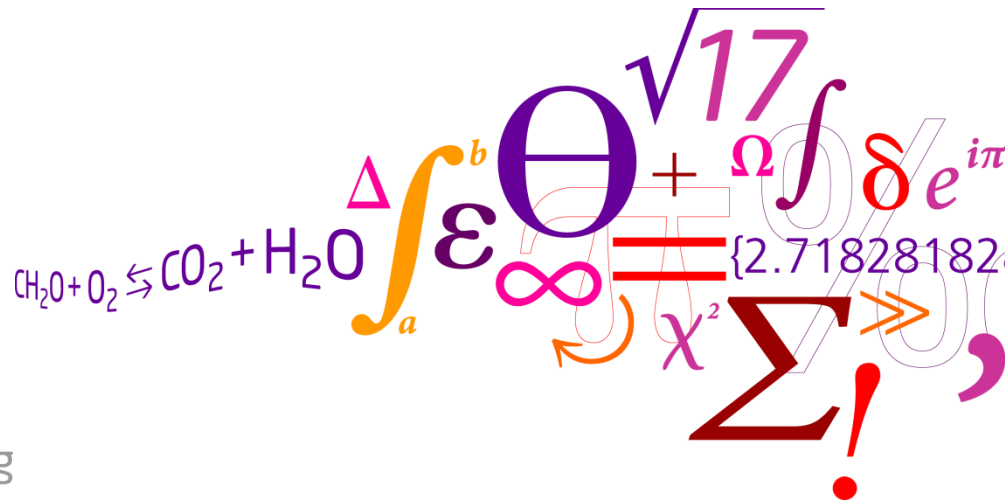
<http://metlab.rt.env.dtu.dk/>

**bfsm@env.dtu.dk**

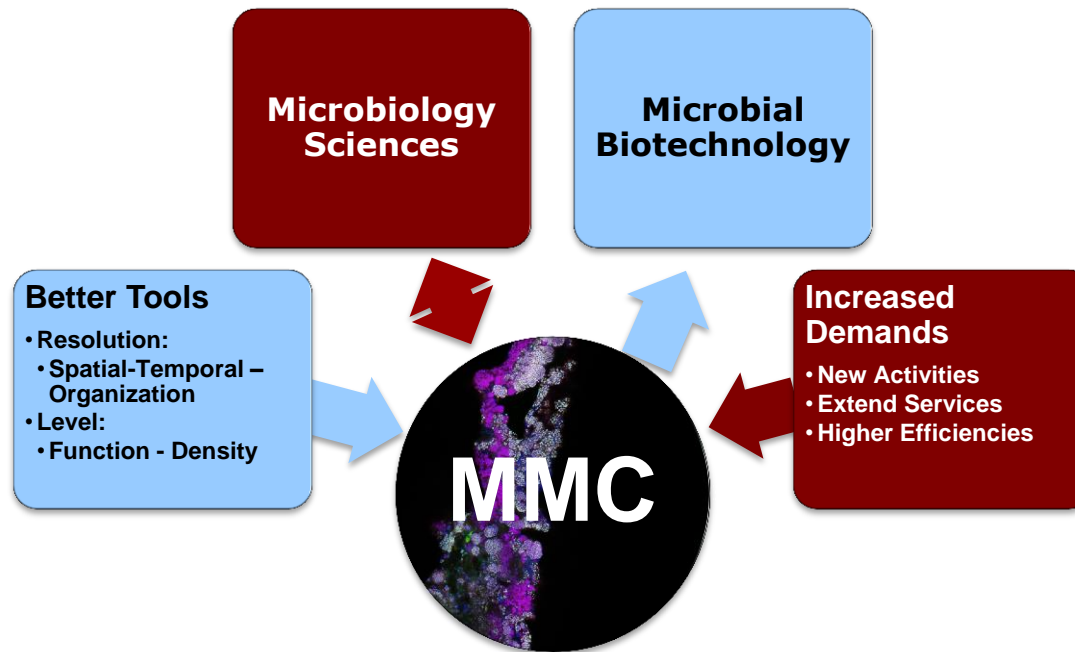
**DTU Environment**

Department of Environmental Engineering

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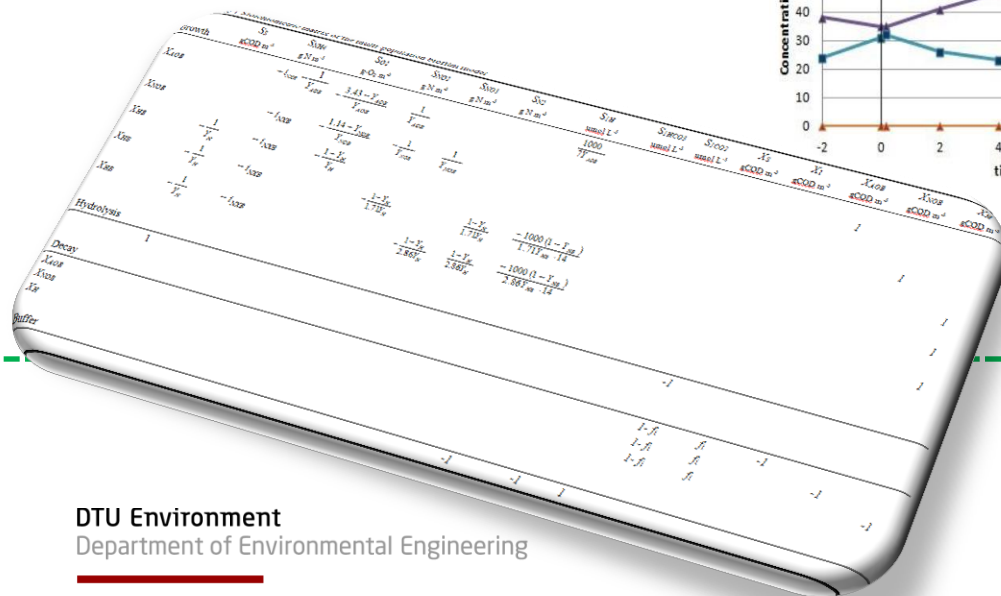
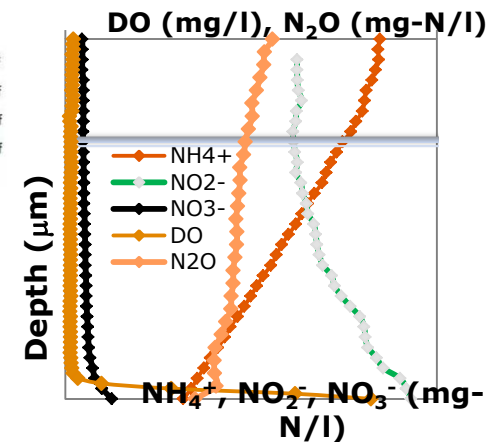
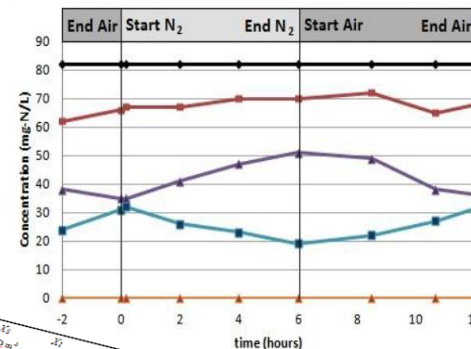
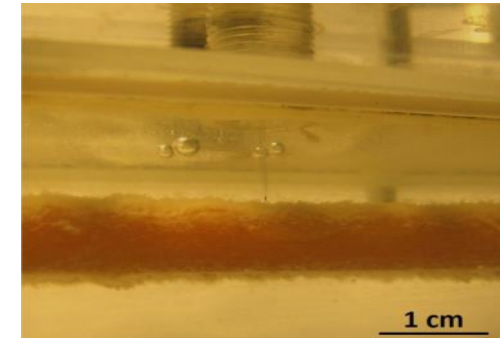
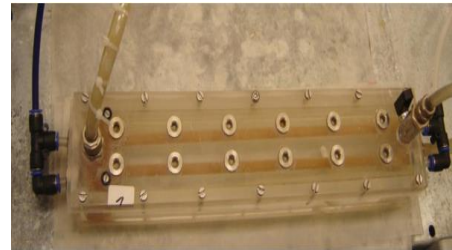
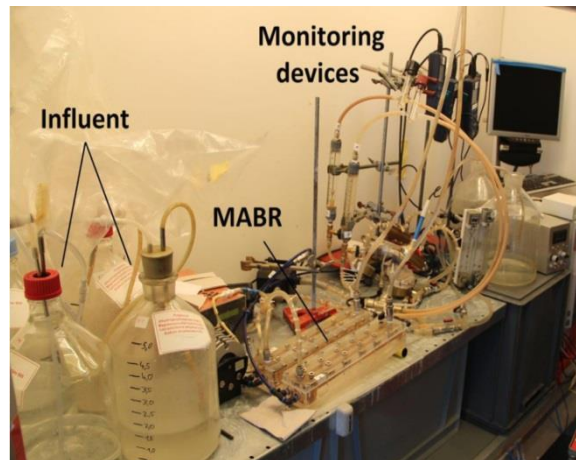
Managing and Engineering Open Microbial Communities to Attain Specific Services for the Use and Benefit of Society and the Environment.

## **MICROBIAL RESOURCE MANAGEMENT & ENGINEERING**

at the interface of microbial ecology and environmental engineering



# Model-based Evaluation of Nitrogen Removal in Membrane-Aerated Biofilm Reactors





# Model-based Evaluation of Nitrogen Removal in Membrane-Aerated Biofilm Reactors

- **Background**

Membrane-aerated biofilm reactor is an excellent candidate to perform autotrophic nitrogen removal, which is both cost- and energy-efficient. Aquasim is a popular modelling program for simulation of wastewater treatment systems, helping us to know more details about these biological processes.

- **Aim**

Operate a counter-diffusion biofilm reactor to realize partial-nitrification N removal process.

Build a multi-species nitrifying biofilm model to evaluate the removal performance under various operational conditions, and identify strategies to suppress unwanted microbial types (Nitrite oxidizing bacteria, NOB).

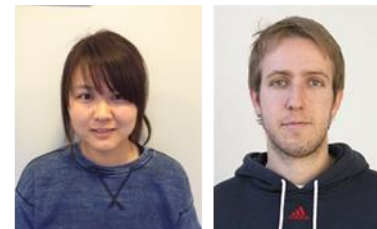
- **What you will learn...**

Experimental: Run lab-scale biofilm reactors, and monitor the daily performance by  $\text{NH}_4^+$ ,  $\text{NO}_2^-$ ,  $\text{NO}_3^-$  and DO measurements. Measure N species/profiles within the biofilm (microsensors).

Computational: A one-dimensional biofilm model (AQUASIM).

**Supervising PhD/PD** – Yunjie Ma

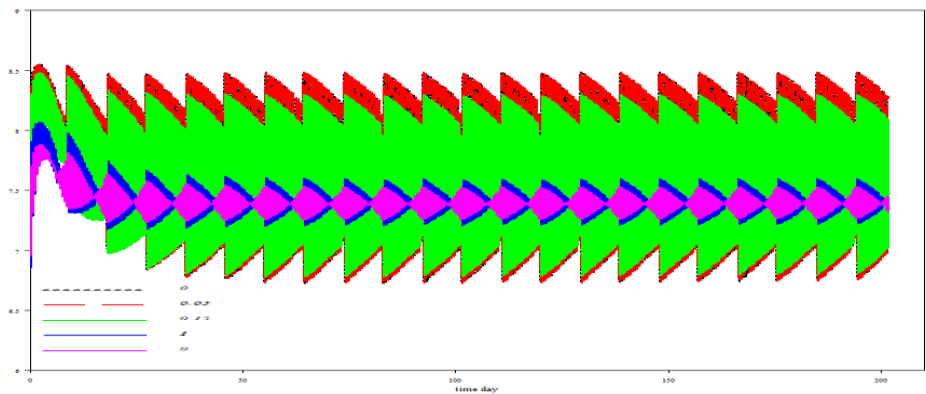
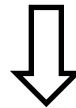
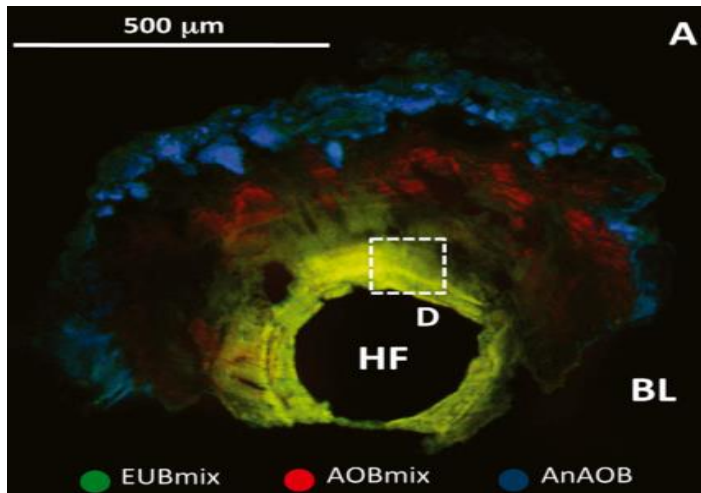
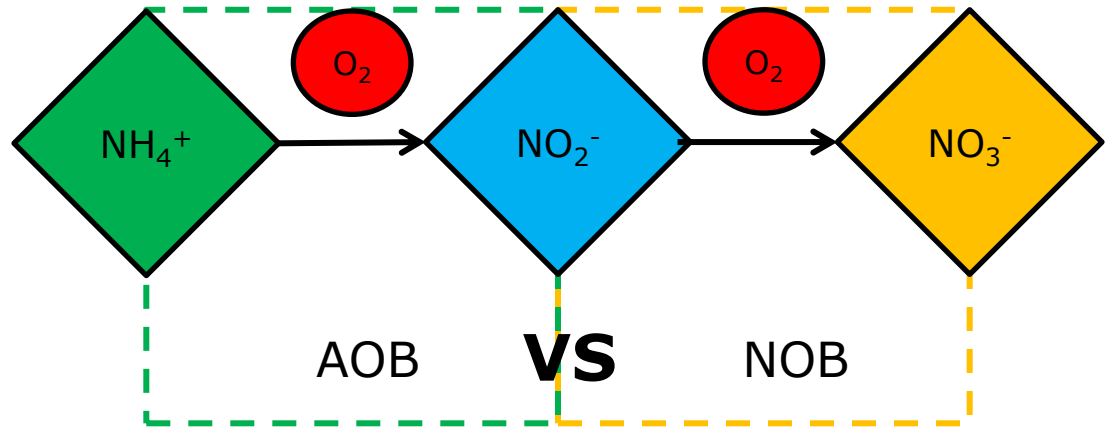
Carlos Domingo Felez





# NOB suppression Mechanism

- Various operational conditions: ASL, DO setpoints, pH control, buffer ratios.
- NOB suppression: DO, FA, FNA, pH.





# NOB suppression Mechanism

- **Background**

The "short-cut" nitrogen removal via nitrite is energy- and cost- efficient. BUT to realize this process, NOB (nitrite oxidizing bacteria) is always the trouble on the way!

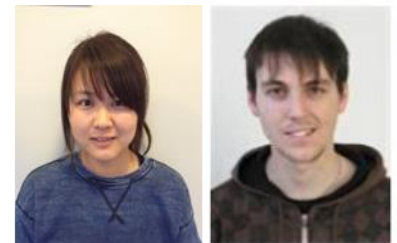
- **Aim**

Evaluate NOB suppression by operating biofilm/suspended sludge reactors under designed experimental conditions, focusing on these 4 factors: dissolved oxygen, free ammonia, free nitrous acid and pH. Elucidate the underlying suppression mechanism...

- **Approach**

Monitor the N removal process, including  $\text{NH}_4^+$ ,  $\text{NO}_2^-$ ,  $\text{NO}_3^-$  daily measurements and microbial community analysis (FISH, qPCR). Design batch experiments to study the relative contributions of the 4 factors mentioned above to NOB suppression. In this work, we will specifically focus on free ammonia limitation and inhibition.

**Supervising** PhD/PhD – Yunjie Ma  
Alex Palomo





# Partial Nitrification in Granule-based SBRs

Monitoring      Reactors

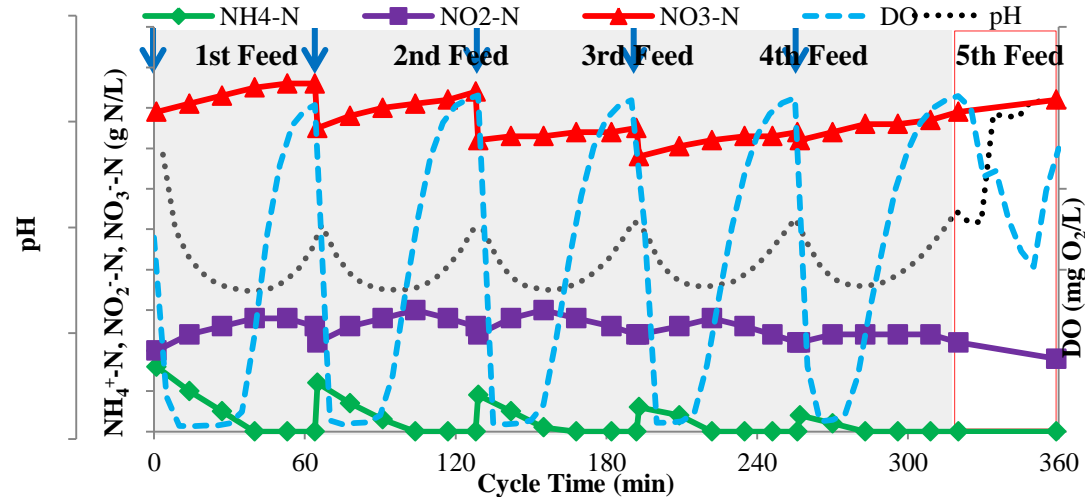
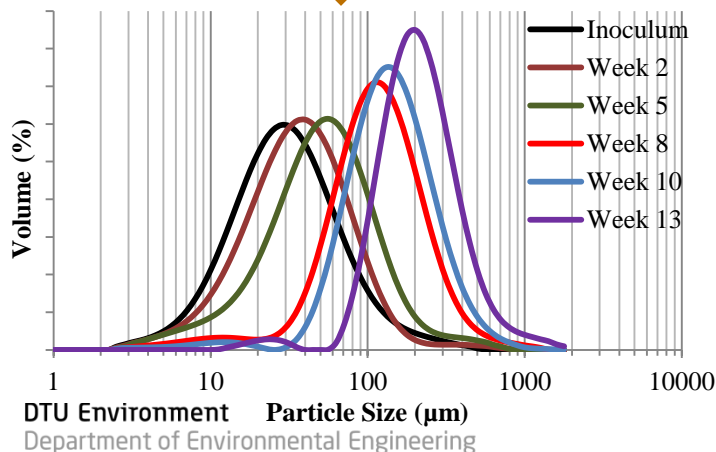
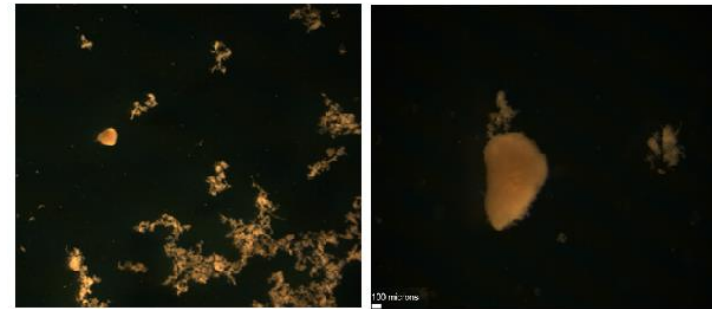
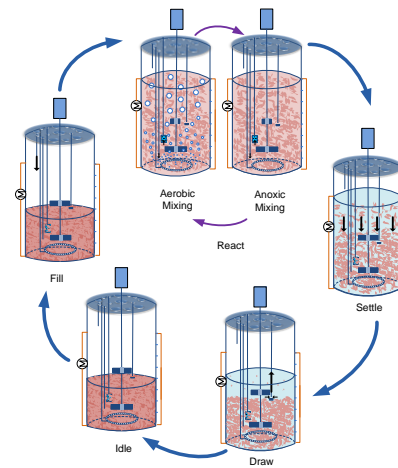


Figure 1. In-cycle analysis of liquid concentrations of a representative SBR<sub>sf</sub> cycle (day 91). The shaded part corresponds to the aerated period, while the blue arrows to feeding times.



# Partial Nitrification in Granule-based SBRs

## • Background

Partial nitrification is an innovative and cost-effective technology for biological nitrogen removal from wastewater.

Nitrous oxide ( $\text{N}_2\text{O}$ ) is a potent greenhouse gas that can be emitted during this process.

## **Saving energy vs. Low GHG emissions**

## • Aim

- Quantify  $\text{N}_2\text{O}$  production kinetics and emissions during partial nitrification
- Identify and quantify the relative importance of the  $\text{N}_2\text{O}$ -producing pathways and associated microorganisms
- Apply operational settings to mitigate  $\text{N}_2\text{O}$  emissions.

- **Approach** - Running bioreactors, monitor performance, microsensors, stable isotopes, molecular analysis.

**Supervising**      PhD/PD Carlos Domingo-Félez  
Marlene M. Jensen





# Who is producing N<sub>2</sub>O, what are they doing?

## • Aim

- Quantify N<sub>2</sub>O production
- Identify N<sub>2</sub>O-producing pathways
- Identify the microorganisms
- Quantify the functional genes

- **Approach** – Experimental, Batch incubations, lab-scale and pilot-scale reactors, stable isotopes, Mass-spec, microsensors, qPCR, pyrosequencing

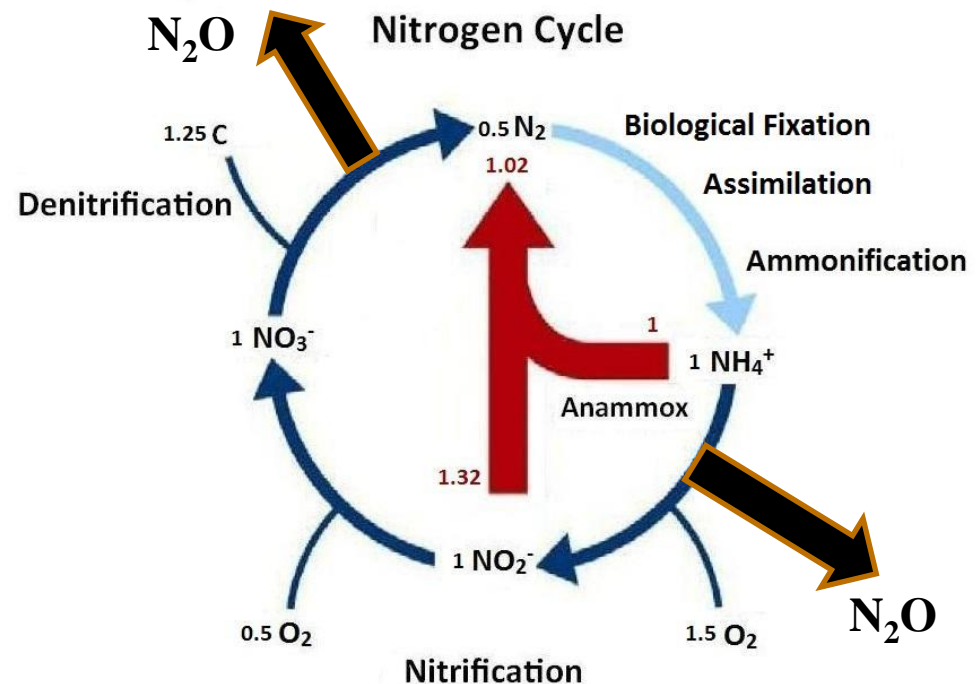
## • Contact

**Marlene M. Jensen**



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Department of Environmental Engineering





# Quantification and modelling of N<sub>2</sub>O emissions from N-removing biomass

## • Background

Nitrous oxide (N<sub>2</sub>O) is a potent greenhouse gas emitted during biological nitrogen removal from wastewater.

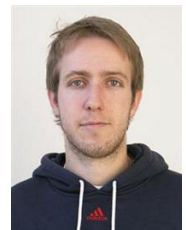
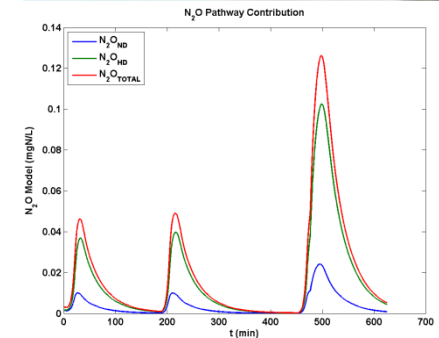
## • Aim

Establish and calibrate a consensus process model to assess N<sub>2</sub>O dynamics

## • Approach

- Modelling: Evaluate current models and/or develop new models to describe N<sub>2</sub>O production from WWTPs.
- Experimental: Develop an assay to estimate N<sub>2</sub>O production with biomasses from different WWTPs.

## • Supervising PhD/PD - Carlos Domingo-Félez





# N<sub>2</sub>O dynamics – Full-scale and pilot-scale measurements

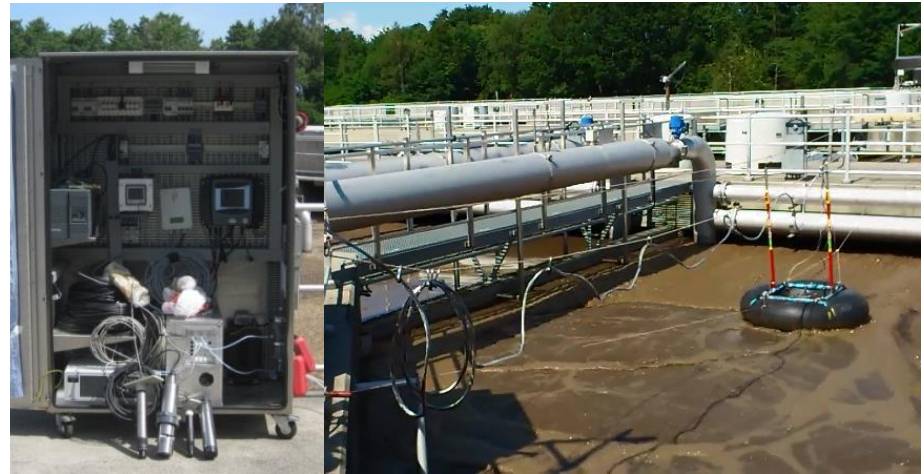
Online data acquisition

Liquid phase:

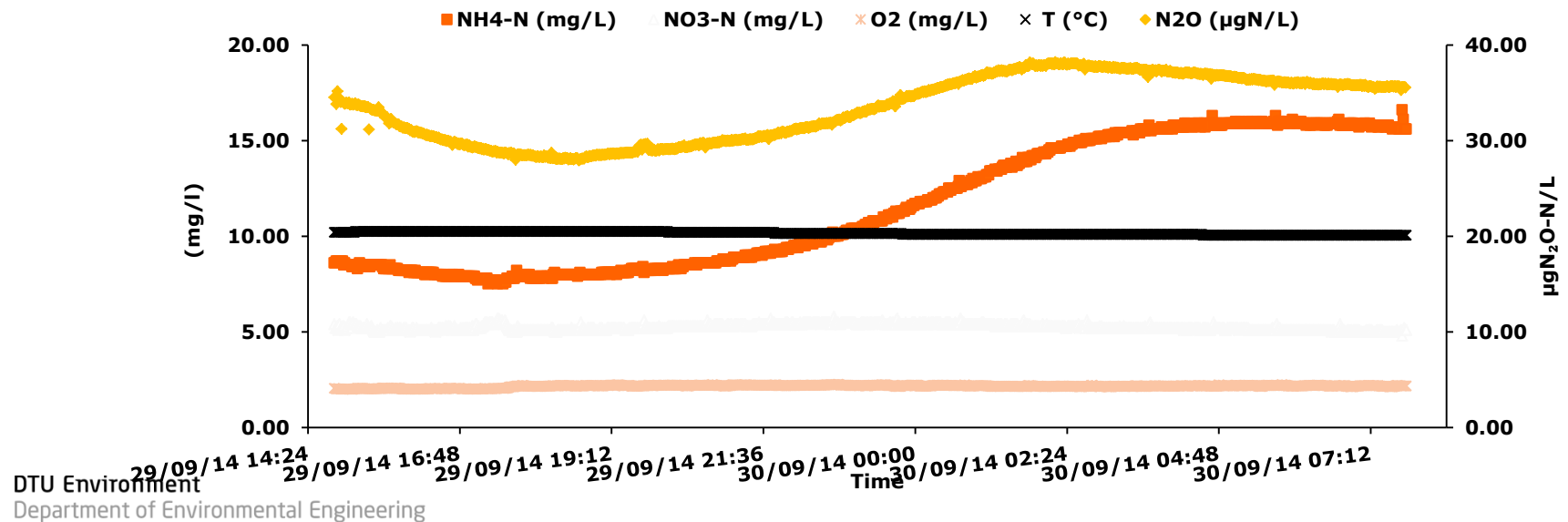
NH<sub>4</sub>-N, NO<sub>3</sub>-N, N<sub>2</sub>O, pH & O<sub>2</sub>

Gaseous phase:

N<sub>2</sub>O, O<sub>2</sub> and CO<sub>2</sub>



Example from Manammox-pilot:





# N<sub>2</sub>O dynamics – Modelling of full-scale observations

## System configuration

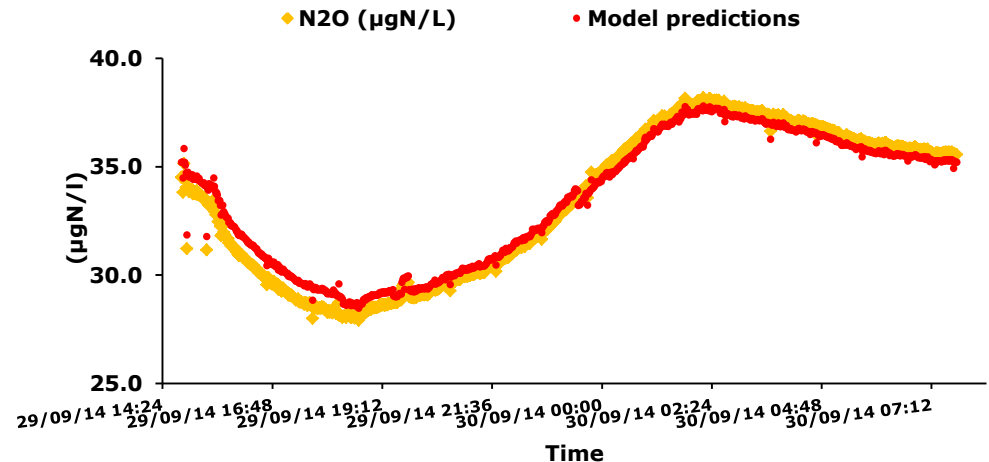
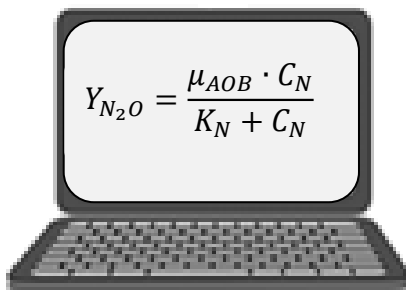
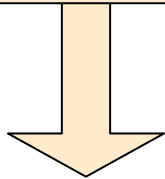
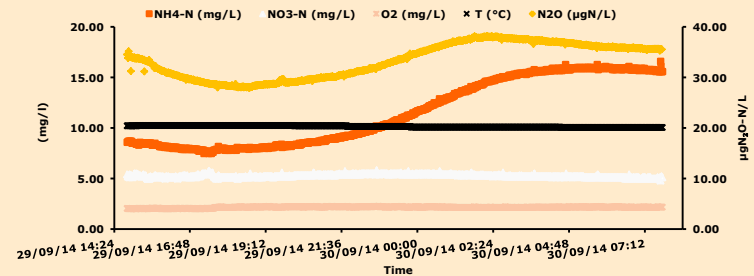


## Stoichiometric matrix for model development

Process	S N <sub>2</sub> O-N	S O <sub>2</sub>	....
1	$Y_{AOB} - 1$	-1.14	....
2	3	$1.14 \cdot Y_{AOB}$	....
3	1	2	....
....	....	....	....

\*Fictive matrix

## Online data

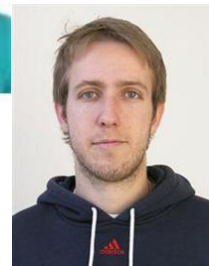




# Full-Scale/Pilot Scale N<sub>2</sub>O Measurements

- **Aim** – Quantify N<sub>2</sub>O dynamics at full-scale and pilot-scale
- **Approach**
  - Experimental: Field measurements, oxygen, CO<sub>2</sub> and bulk N species with biosensors, microsensors, GC
  - Computational: Process Modelling to full-scale N<sub>2</sub>O observations

- Supervising PhD/PD – **Sara Ekström**  
**Carlos Domingo-Félez**  
**Marlene M. Jensen**

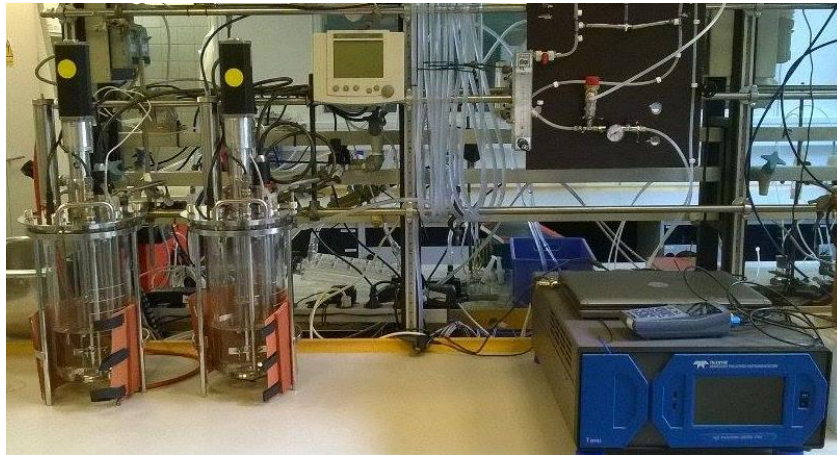


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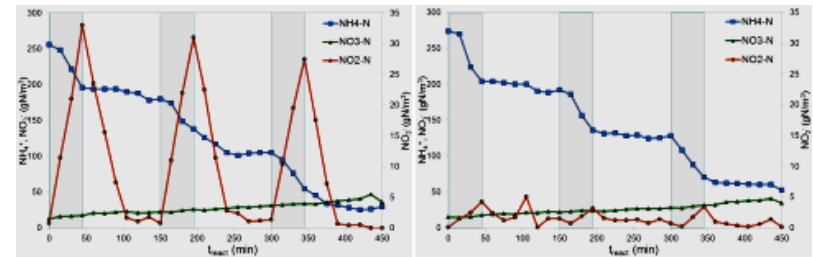


# Understanding Partial Nitrification/Anammox to avoid $N_2O$ emissions

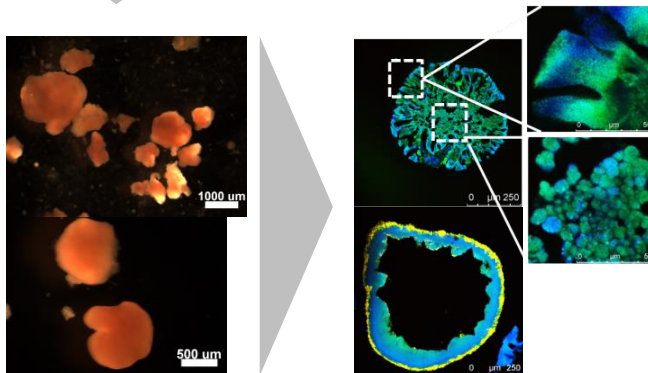
1. Cultivate AOB/AMX in LabScale



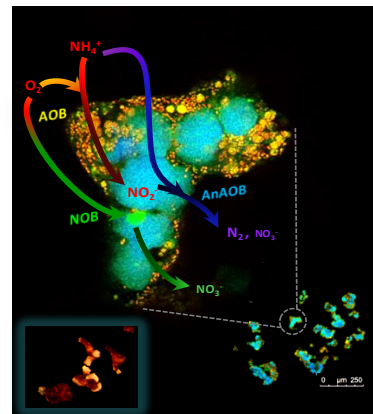
5. Find a strategy to mitigate  $N_2O$  emission from PN/A



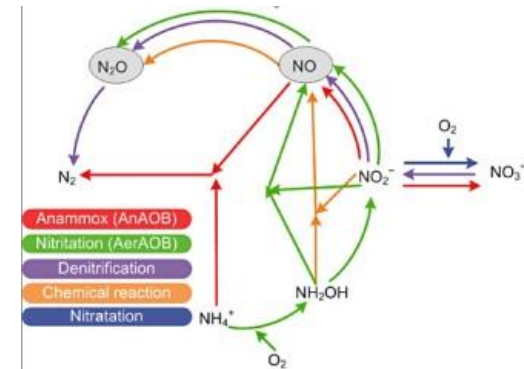
2. Study microbial community in granules



3. Understand processes and interactions



4. Identify and engineer an "ideal" microbial community





# Linking N<sub>2</sub>O emissions to microbial community structure

## ● Background

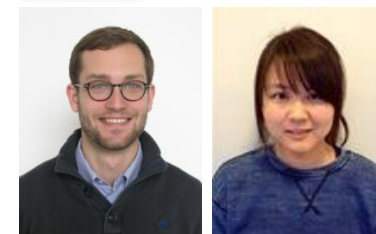
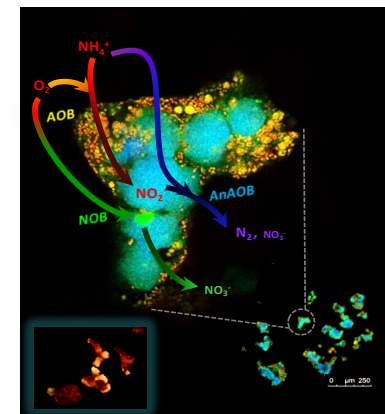
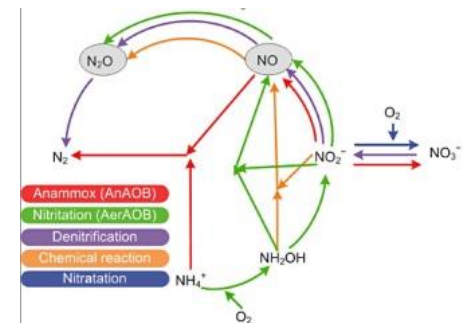
Biological nitrogen removal from wastewater is performed by the partial nitrification-anammox process. However, emissions of the greenhouse gas nitrous oxide (N<sub>2</sub>O) from the process are of concern and mitigation strategies of N<sub>2</sub>O emissions are needed. The sinks and sources of N<sub>2</sub>O in the metabolism of involved microorganisms are in the spotlight to achieve a sustainable nitrogen removal process.

## ● Aim

The project will elucidate a potential relationship between the particle size of biogranules, their inherent microbial community and N<sub>2</sub>O emissions. The findings will help to judge whether certain granules have superior properties over others and should be applied to the process preferentially.

## ● What you will learn

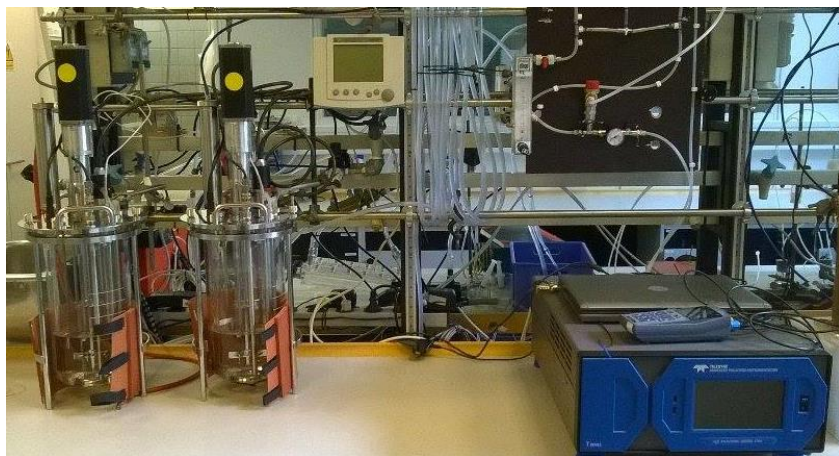
Microbial community analysis (FISH, qPCR), reactor handling, process monitoring, data handling and interpretation, fluorescence microscopy



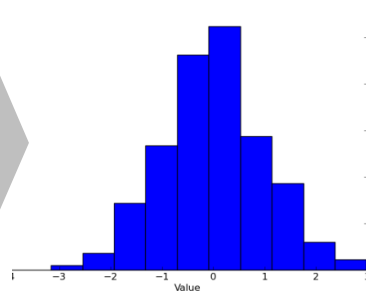
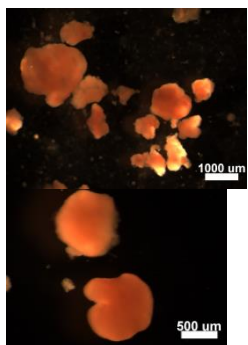


# Linking N<sub>2</sub>O emissions to microbial community structure

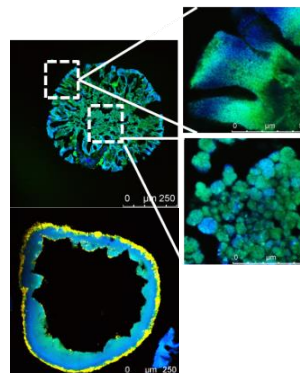
## 1. Cultivate AOB/AMX in LabScale



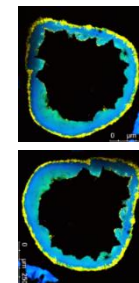
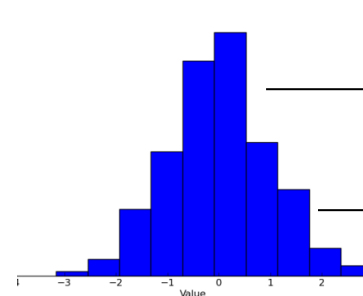
## 2. Identify particle size distribution



## 3. Study microbial community in granules



## 5. Analyze N<sub>2</sub>O emission as f (particle diameter)



x N<sub>2</sub>O

y N<sub>2</sub>O

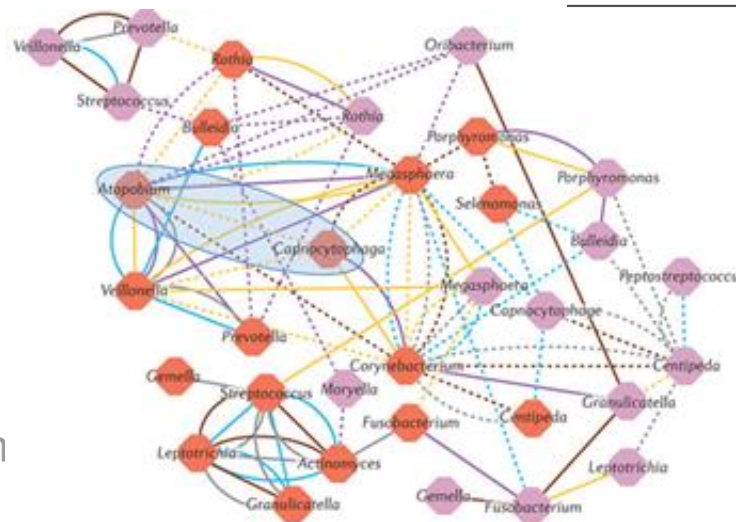
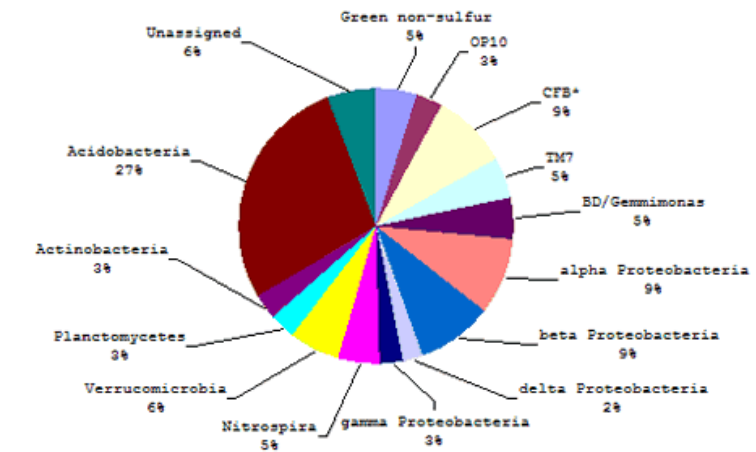
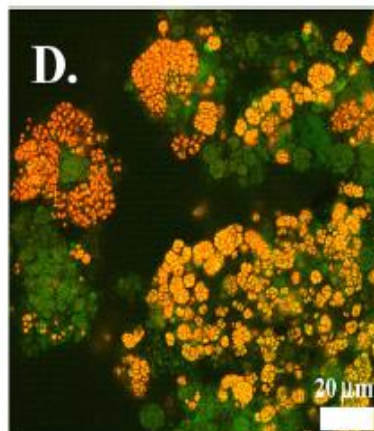
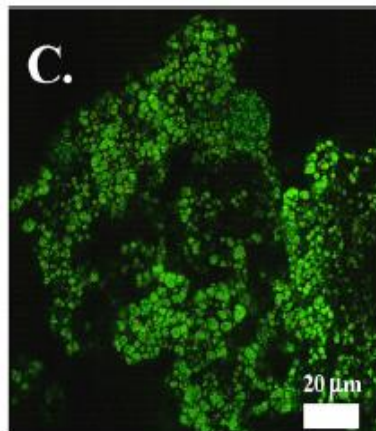
## 4. Analyze N<sub>2</sub>O emission of different granule sizes

N<sub>2</sub>O





# Systems microbiology to understand microbial communities



$$\epsilon \Theta^{v17} + \Omega \int \delta e^{i\pi} = \{2.71828182\} \chi^2 \Sigma!$$



# Community assembly processes in guilds of nitrite oxidizing bacteria

## • Background

Community assembly processes can be used to explain microbial community's response to, for example, outside stress, resource pulse and invaders. Ecologists consider the four main community assembly processes – selection, dispersal, drift and speciation – as the key to understanding ecological interactions between species, guilds and communities.

## • Aim

- Characterize the guild composition of nitrite oxidizing bacteria (NOB) in engineered communities
- Study the extent of different community assembly processes – selection and/or dispersal – in NOB guilds

## • What you will learn

- Operate bioreactors under defined conditions to enrich for NOB dominated microbial biofilms.
- Monitor the microbial activity in the bioreactors and use different molecular methods to characterize microbial community composition
- Evaluate the effect of selection and/or dispersal in shaping that guild composition

**Supervising** PhD Marta Kinnunen





# Which microorganisms are treating the water we drink?

- **Background:**

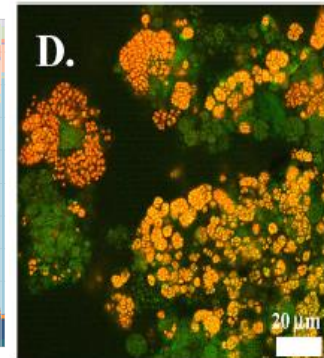
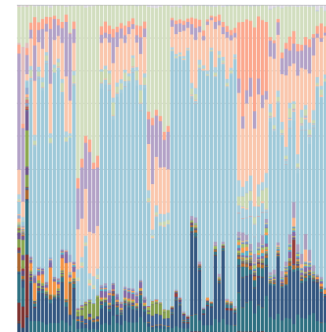
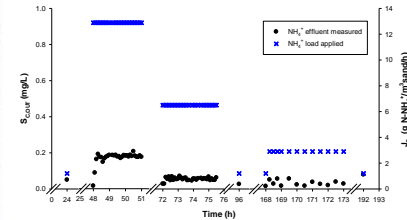
Rapid sand filtration is a widely used drinking water production technology. This technique allows the removal of pollutants from groundwater.

- **Objective:**

Operate a lab scale biofilter under different conditions and identify the microorganisms involved in the removal of several compounds.

- **What will you learn:**

Reactor handling, process monitoring and data interpretation, microbial community analysis (FISH, qPCR).



**Supervising PhD/PD** – Alex Palomo  
Jane Fowler

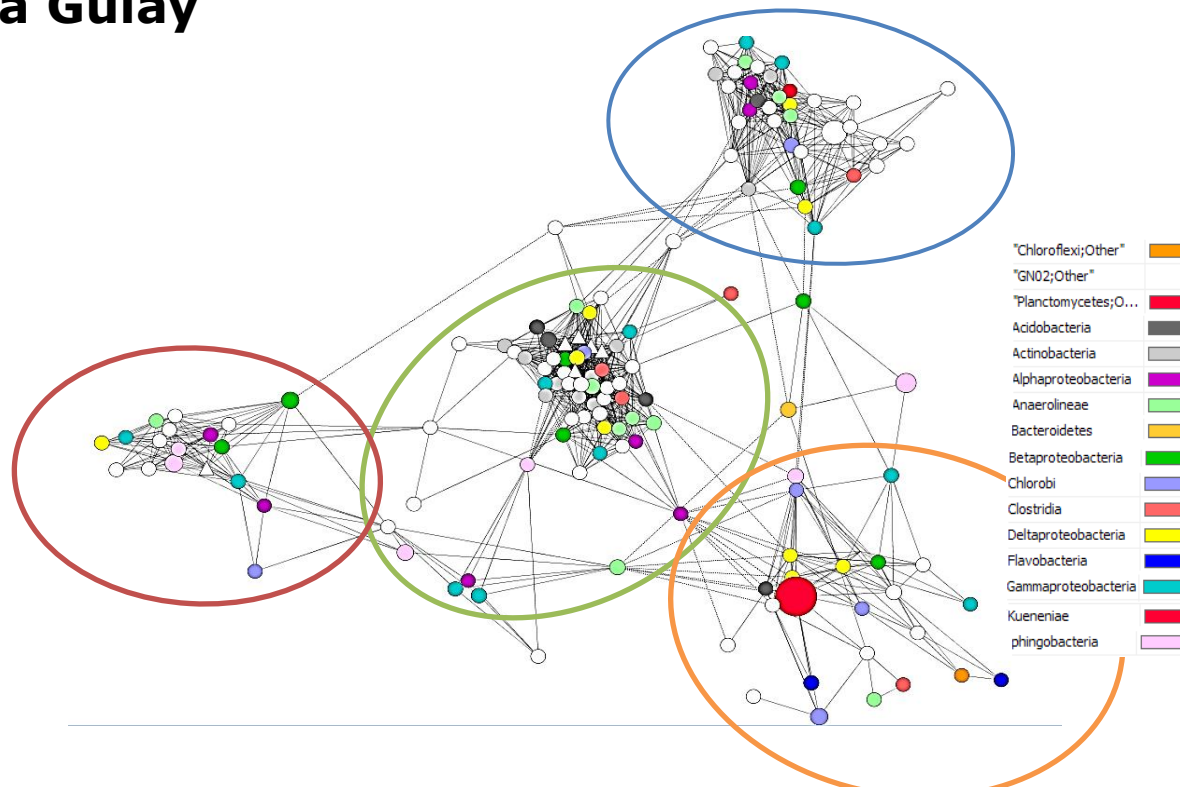




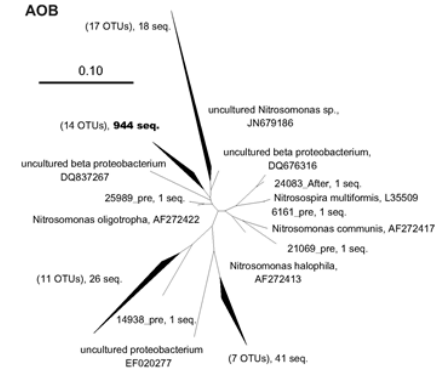
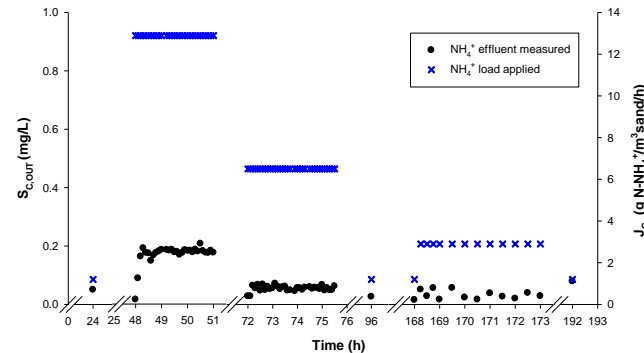
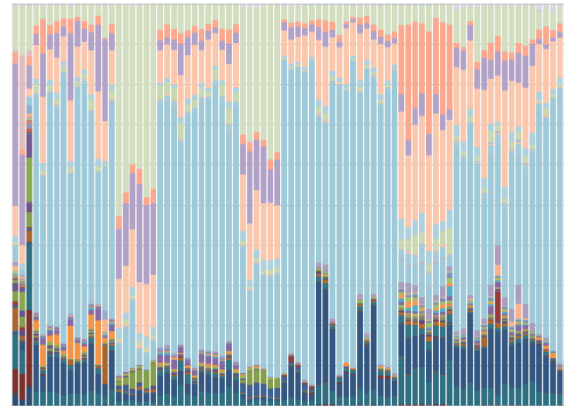
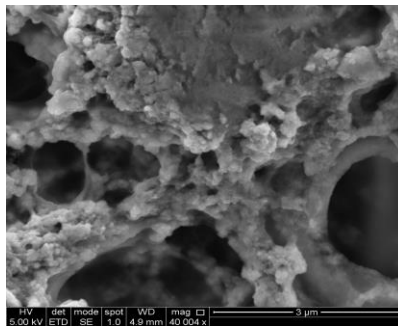
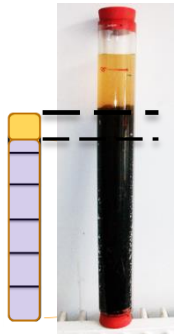
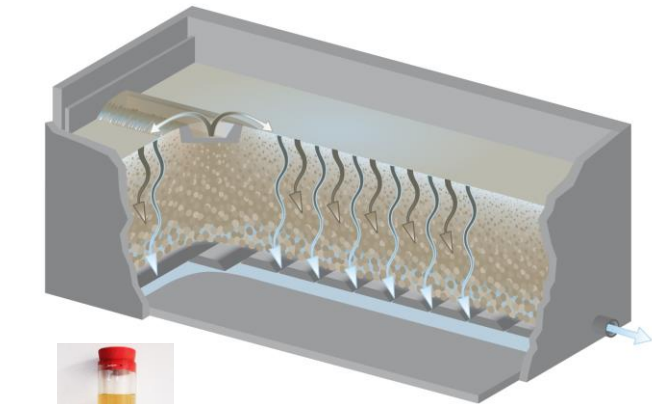
- **Aim** – study the diversity, networks, and expression of genes in WWTPs
- **Approach** - Omics
- Supervising PhD/PD – **Alex Arda Gülay**



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argl@env.dtu.dk

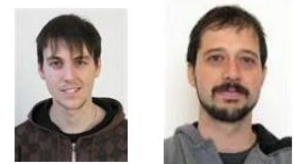






**FOR PROJECTS IN THE AREA OF WATER TREATMENT BIOTECHNOLOGY –  
ELUCIDATING THE COMPOSITION, FUNCTION, AND RATES OF MICROBIAL  
COMMUNITIES IN WATER FILTRATION PROCESSES**

**SEE PHD STUDENTS ALEX PALOMO, PD ARDA GULAY**





# Computational study of microbial community dynamics

## • Background

Models are useful tools to gain a better understanding of a 'system' and to test hypotheses about this system. In the field of microbial ecology, computational models are increasingly used to study complex communities and processes such as competition and coexistence in multispecies biofilms.

## • Aim

Use and extend existing microbial modelling tools (iDynoMiCS) to study competition, coexistence and cooperation in microbial aggregates. Identify key factors that lead to specific individual 'behavior' and assess how this affects the community fitness.

## • What you will learn

You will learn how to use established software to answer your scientific questions. You will contribute by developing extensions to this software in order to introduce novel mechanics to your own model. (Affinity with programming is recommended for this project)

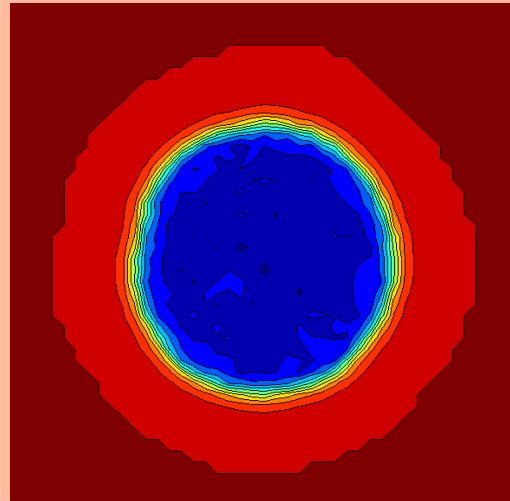
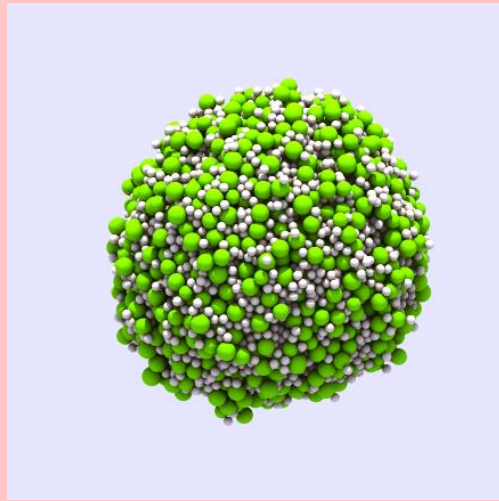
**Supervising**

PhD - Bastiaan Cockx

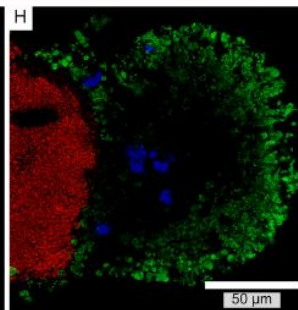
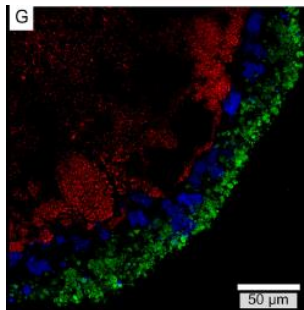
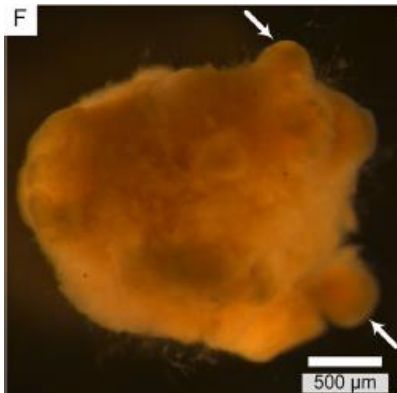
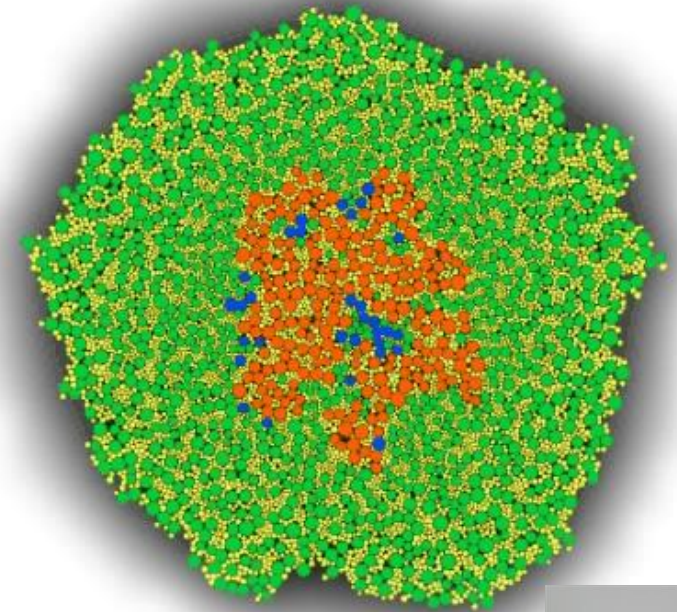




# Computational study of microbial community dynamics



iDynoMiCS



Vlaeminck et al. 2010

**Supervising**

PhD - Bastiaan Cockx





# Current Research Team

## Core

- **Dr. Arnaud Dechesne**
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